

Cortical organization: neuroanatomical approaches

Study of neuronal circuitry
and microstructure

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Changes in characteristics
seen through phylogeny

Increase in the absolute and particularly
relative mass of the brain compared to body size

Comparatively larger increase in telencephalic
structures particularly the cerebral cortex

Expansion in number of radial units

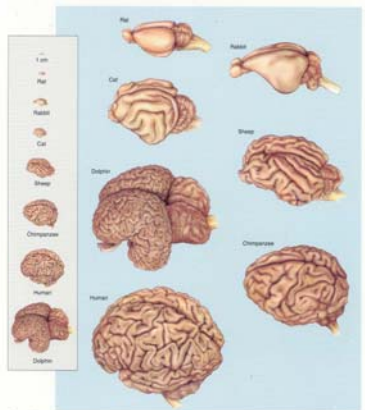
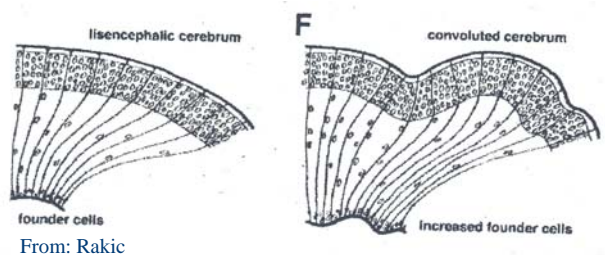


Figure 7.1
Mammalian brains. Despite differences in complexity, the brains of all these species have many features in common. The brains have been drawn to appear approximately the same size; their relative sizes are shown in the inset to the left.



Neoteny: prolongation of fetal growth rate into postnatal times results in increase in the size of the human brain

Evolution of the Nervous System

How does a larger or reorganized brain come about?

Environmental factors

Internal factors – change in posture

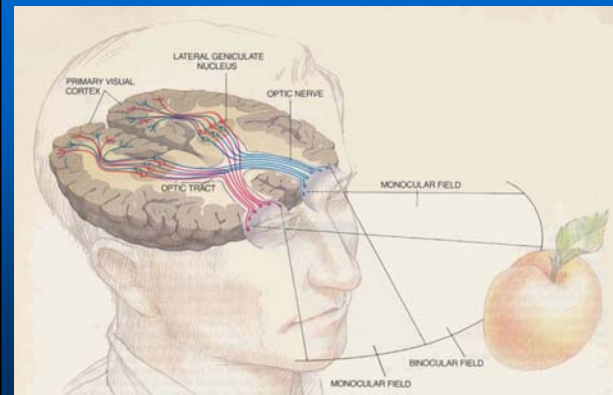
freedom of the hands

movement of eyes from a lateral to central position

Crossing at the optic chiasm is complete in animals with small degree of binocular vision

FREEDOM OF THE HANDS

Direct projection to the pyramidal tract in animals with fine use of the hands



How did these changes come about in evolution?

By what mechanism?

Distinct modes of radial migration:

Inside-out: characterizes pattern in mammalian cortex

Outside-in: characterizes reptilian cortex

Consequences of differential migration: differences in cortical thickness, synaptic interactions

reelin (glycoprotein)

mice deficient in reelin have inverted cortical lamination

cdk5 (cyclin-dependent protein kinase)

mice lacking cdk5 or its activator p35 show inverted cortical lamination

Differences in the two mutants:

in cdk5/p35 mutants the marginal zone and subplate are properly differentiated;

in the reeler they are not

Addition of new cells to the primate CNS

Example from humans

Spindle cells in cingulate gyrus

GABAergic cells in the dorsal thalamus (DT)

Large expansion of 'association' areas

Progressive change from the general to the specific; loss of some connections

Differentiation of cortical layers

Structures that are large relative to overall brain size have late birthdays

Why?

Building the Brain

Brainstem

Thalamus and hypothalamus

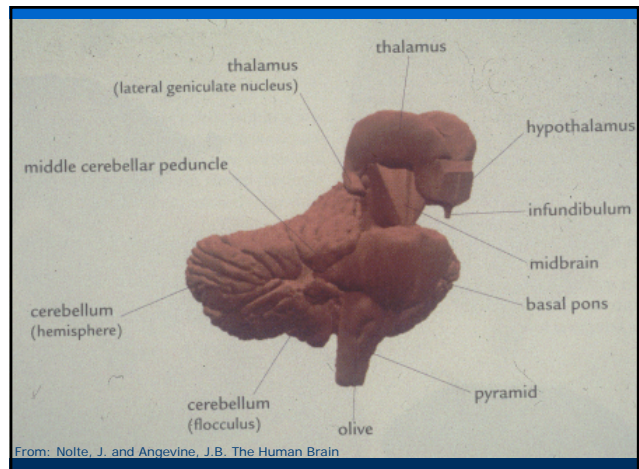
Ventricles

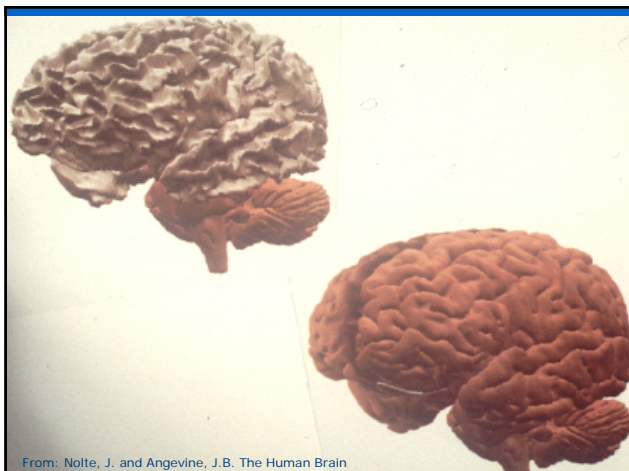
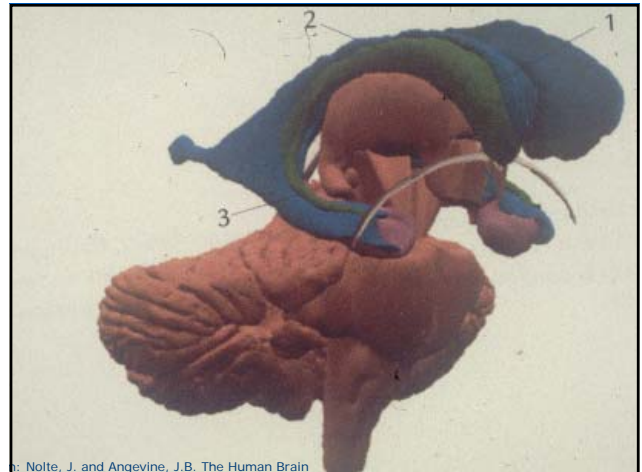
Basal Ganglia

Amygdala

Hippocampus

Pathways





MAJOR INFLUENCES ON CORTEX

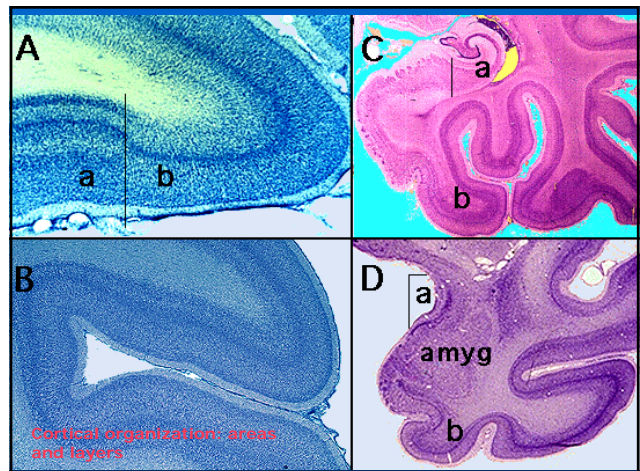
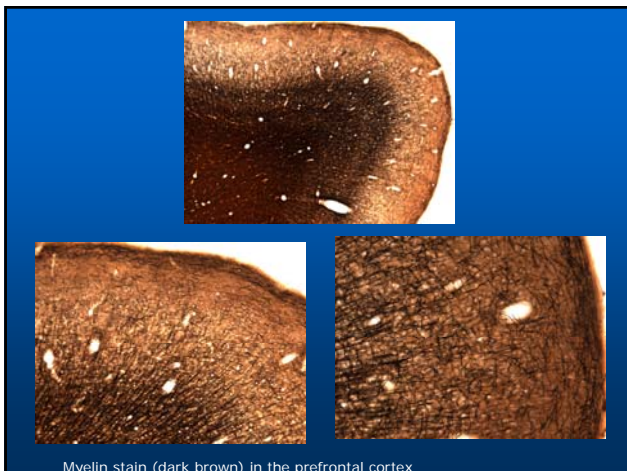
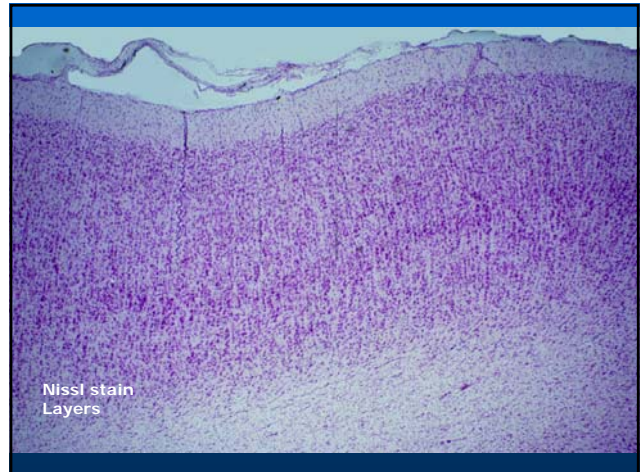
Neurotransmitter- specific
projection systems: Cholinergic;
Adrenergic; serotonergic; Dopaminergic
arise from subcortical structures

Thalamus/hypothalamus

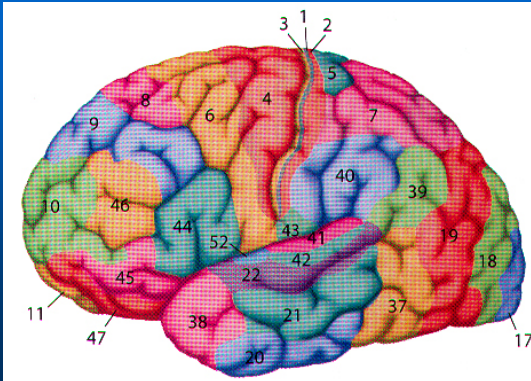
Specialized projections:
Hippocampal formation
Amygdala

Methods to study cortical architecture

Global structure: classical methods
Nissl stain (stains all neurons
and glia)
myelin stain



Cerebral Cortex: Brodmann map



Classical stains:

Nissl stain
myelin

Are these methods useful today?

Applications

stereological analysis to estimate
density or overall number of
neurons

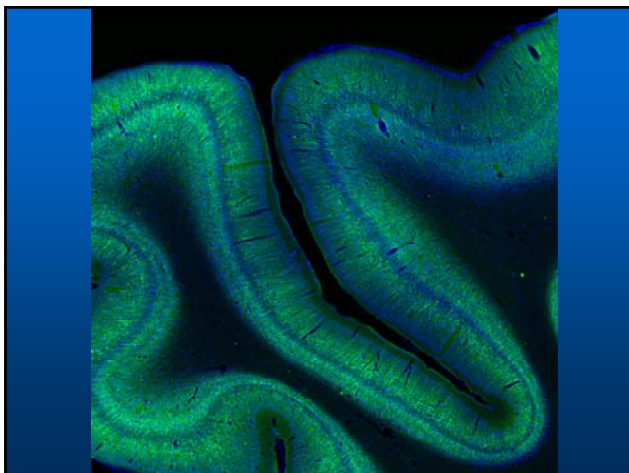
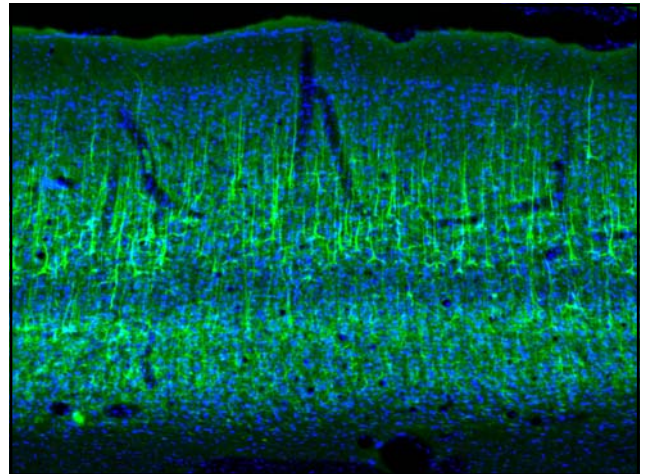
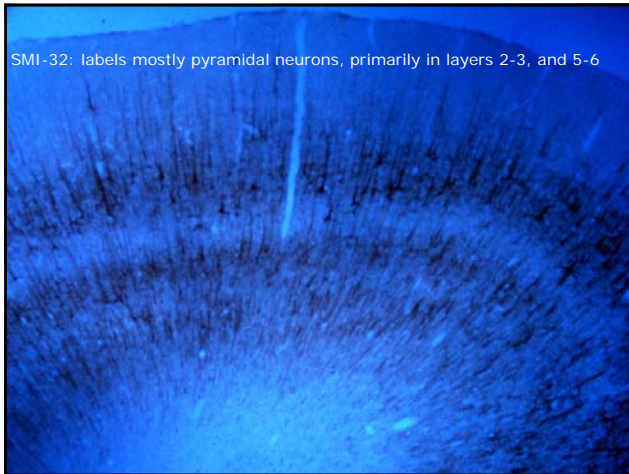
Myelinogenesis: The method was introduced by Flechsig; it makes use of the fact that different fiber tracts become myelinated at different times in their development. Thus, study of the nervous system in embryos and in early neonatal life often provides information about the existence and locality of the different fiber tracts.

Myelination occurs throughout development, and up to the fifth decade of life, and can be used to show development of pathways, particularly in children and adolescents.

Selective labeling of classes of neurons

Histochemistry

Immunohistochemistry



MAJOR CORTICAL CELL TYPES:

Principal or Projection neurons:

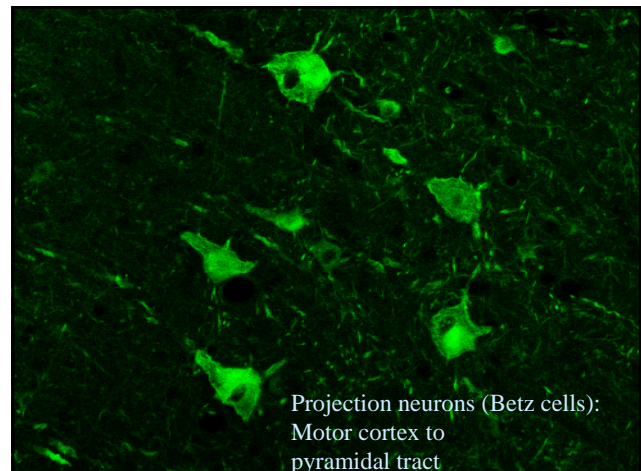
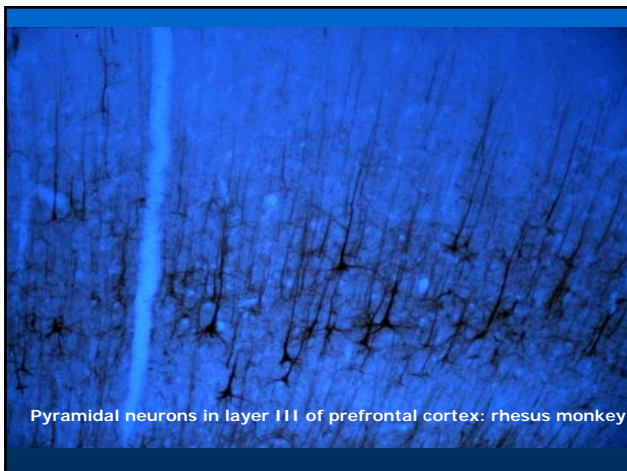
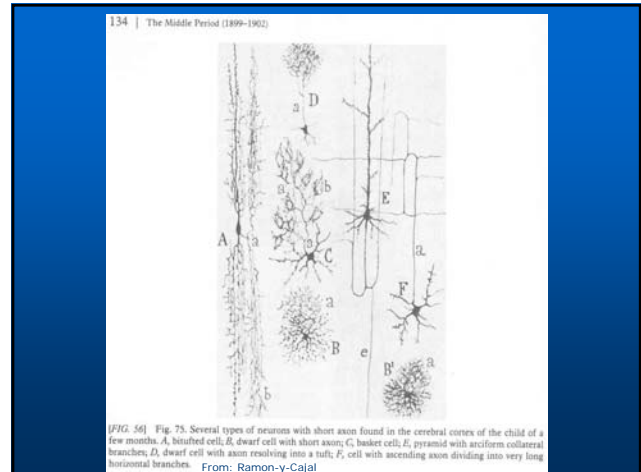
Pyramidal: project outside of immediate area to other cortices or to subcortical structures

Other shapes (e.g., fusiform)

Interneurons:

Excitatory: stellate: small local neurons

Inhibitory: (many different shapes)



Neurochemical markers selectively label distinct classes of neurons in the brain:

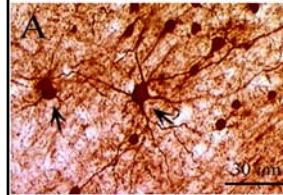
Labeling of inhibitory neurons in the brain

e.g., GABA (GAD)

Neurochemical classes of inhibitory neurons labeled with the calcium binding proteins: parvalbumin; calbindin; calretinin (label inhibitory neurons in the cortex and amygdala but not in thalamus)

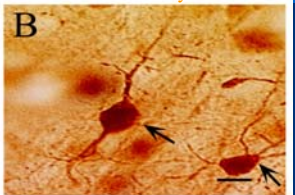
Differences in neurochemical classes of inhibitory interneurons

Parvalbumin inhibitory neurons



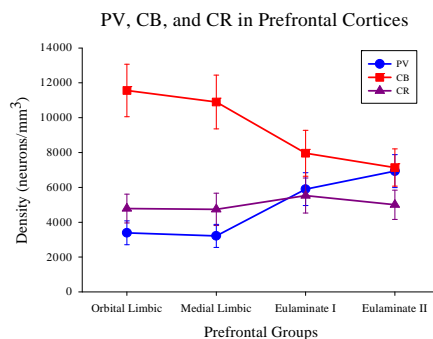
Parvalbumin positive neurons predominate in the middle cortical layers; they are basket or chandelier type inhibitory neurons, targeting the proximal dendrite or axon initial segment of other neurons.

Calbindin inhibitory neurons



Calbindin positive neurons predominate in the superficial cortical layers; they are double bouquet type inhibitory neurons, targeting the distal dendrite of other neurons.

Regional distribution of neurochemical classes of inhibitory neurons



Methods to study cellular architecture

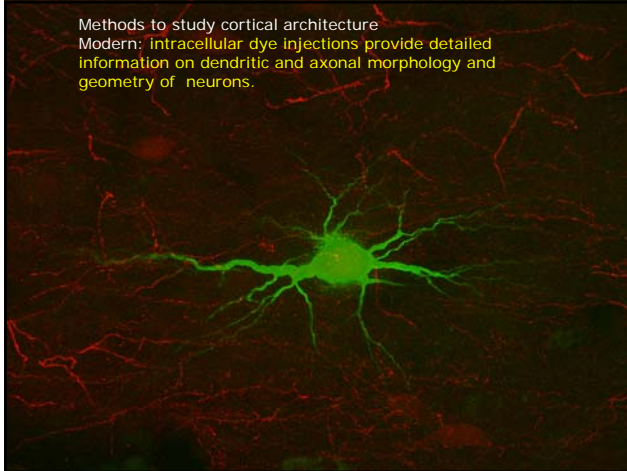
Classical:

Single cell structure: Golgi method

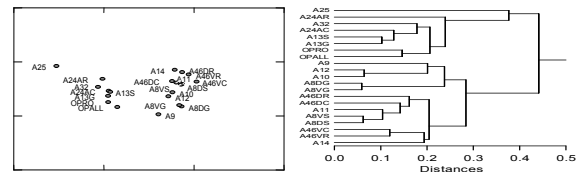
The Golgi method provides detailed information on dendritic and axonal morphology and geometry of neurons.

Drawback of method: labeling is random and unpredictable

Methods to study cortical architecture
Modern: intracellular dye injections provide detailed information on dendritic and axonal morphology and geometry of neurons.



What do architectonic methods tell us about cortical organization?



Methods to study connections

Classical:

Anterograde degeneration: After a lesion axon terminals and the surrounding myelin undergo degeneration and degenerating axons and terminals can be stained selectively.

Retrograde degeneration: After a lesion cell bodies that lose their postsynaptic input degenerate: e.g., LGN neurons degenerate after lesion of V1 (in some species).

Classical:

Efferent connections: degenerating axons

After a lesion has been produced in animals or humans and sufficient time has elapsed for anterograde degeneration to set in, the brain can be studied, and degenerated tracts can be localized by one of the following methods:

•Methods that stain degenerating axons (Nauta-Gygax, Fink-Heimer, De Olmos): Silver impregnation techniques that stain degenerating axons and pre-terminals (Nauta-Gygax) or terminals (Fink-Heimer, De Olmos).

Modern methods to study connections

Incorporation of tracers into neurons: Passive and active processes (uptake, transport, etc.). Anterograde, retrograde, and transneuronal transport of the tracers can take place depending on the tracer and methods.

Modern methods of tract tracing:

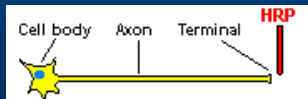
Retrograde tracers: used to study the entire input to an area injected with a tracer.

This method replaced the old retrograde degeneration method, which had significant disadvantages.

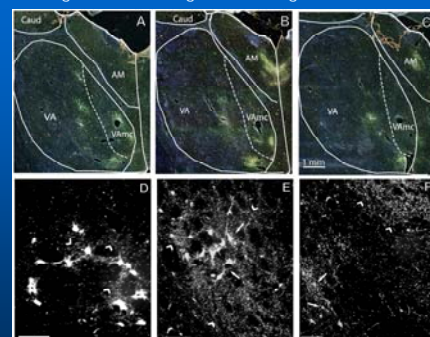
Some retrograde tracers: fluorescent dyes (e.g., fast blue, diamidino yellow); Fluoro-Gold; Cholera toxin, β subunit

•Enzymatic method: When the enzyme horseradish peroxidase (HRP) is injected at the site of termination of nerve fibers, it is taken up by the nerve terminals and transported retrogradely to the perikaryon where it is visualized by an enzyme histochemical technique as blue or brown granules in the soma and dendrites.

•HRP reacts with its substrate, hydrogen peroxide, in the presence of an electron donor (i.e. diaminobenzidine, or tetramethyl benzidine) to yield a brown or blue reaction product, seen by light microscopy; or reacted with osmium tetroxide to yield an electron-dense marker for ultrastructural studies. The HRP marker is widely used for tracing neuroanatomical pathways in the brain.



Bidirectional tracers: HRP-WGA, showing both anterograde and retrograde labeling



From: Xiao and Barbas, Thal. and Related Systems, 2004.

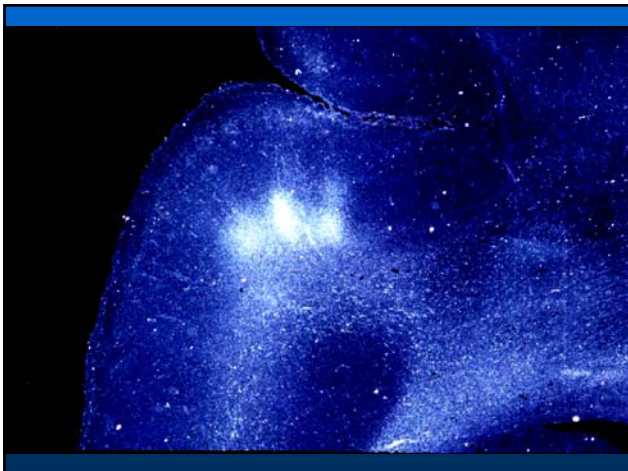
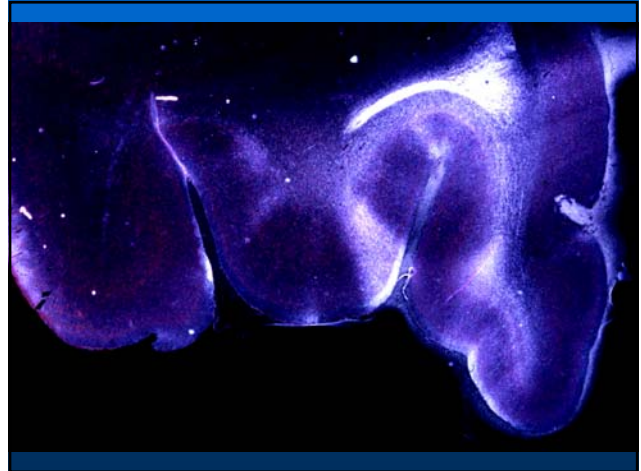
- Modern anterograde tracing (efferent connections)

- Autoradiography:** A method introduced in the 70s, and based on the principle that radioactive amino acids injected in the vicinity of neurons are taken up by the neurons, incorporated into macromolecules, and transported from the cell body down the axon (anterograde transport) to the axon terminal.

- After a finite time following injection, the radioactive amino acid can be demonstrated by autoradiography. By this method, the path of a neural tract can be traced from its origin to its termination.

- This method replaced the old ablation-degeneration methods (Nauta, Fink-Heimer, etc).

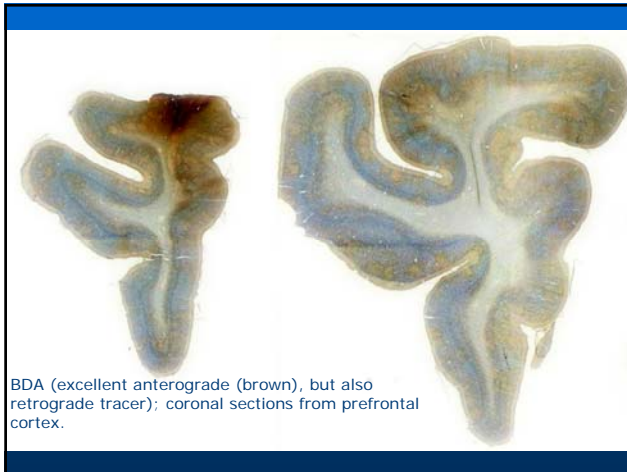
- Some anterograde tracers: ^3H amino acids; BDA



- Bidirectional tracers

- Dextran amines are bidirectional tracers:** Introduced in the late 1980s, the method is highly sensitive, and widely used for anterograde and retrograde pathway tracing studies of the nervous system.

- Dextran amines can be reliably delivered into the nervous system by iontophoresis or pressure injection and visualized with fluorophores or an avidin-biotinylated HRP (ABC) procedure, followed by a standard or metal-enhanced diaminobenzidine (DAB) reaction.



Anterograde and retrograde tracers that work in fixed tissue:

Carbocyanines (DiI, DiO, DiA) for tracing studies in fixed brain tissue

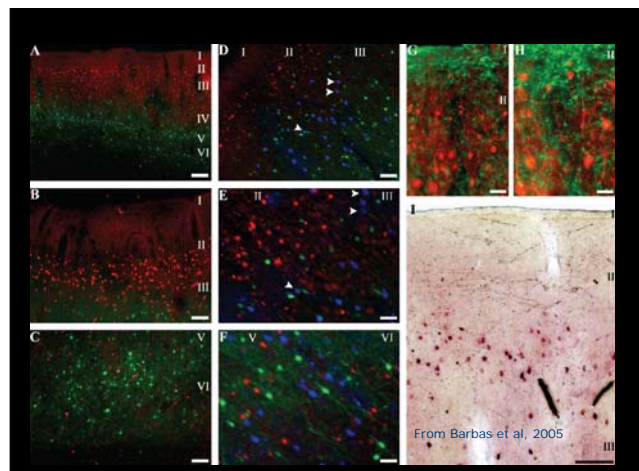
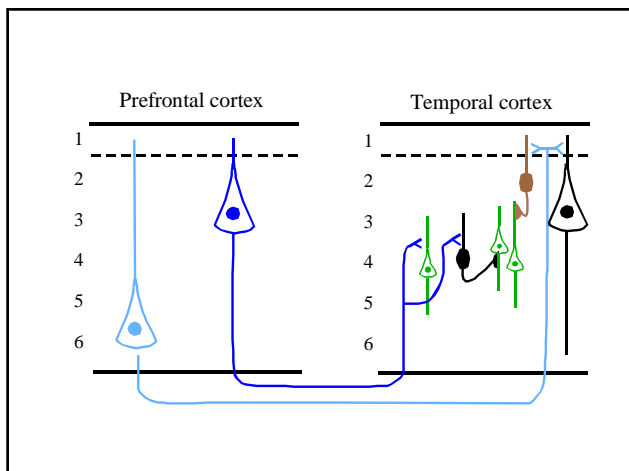
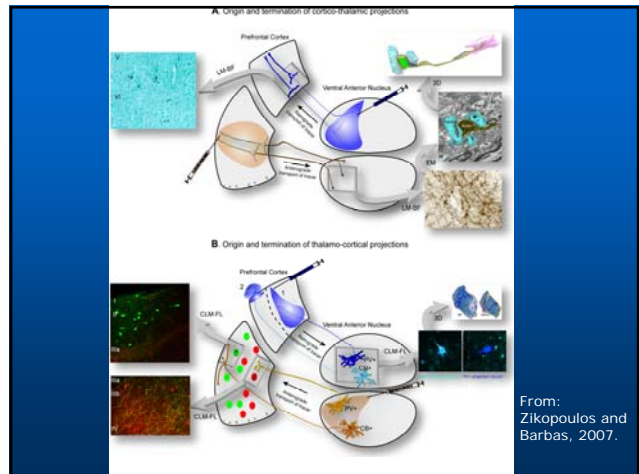
Disadvantage: very long periods are required, especially for human tissue.

•Trans-synaptic transport

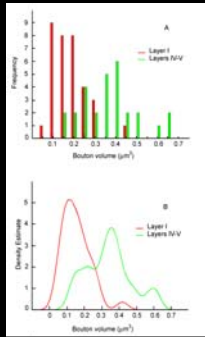
•Anterograde tracers injected in the eye (e.g., ^3H amino acids, HRP-WGA; these dyes work only in this system and were used to show the ocular dominance columns in the primary visual cortex (V1) after injection in the eye.

•Neurotropic viruses for the study of chains of linked neurons; transsynaptic tracing in the retrograde direction (pseudorabies virus)

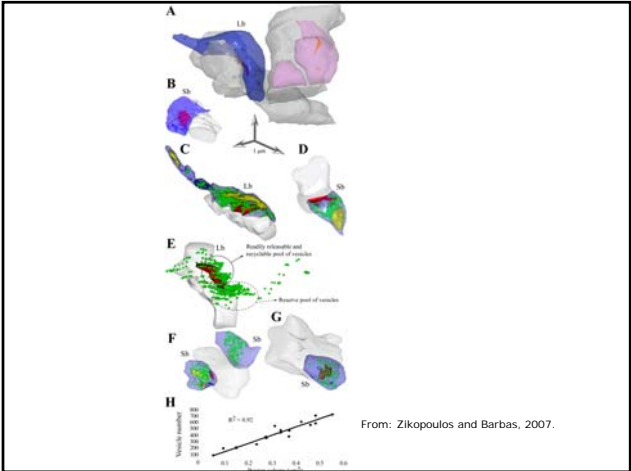
Combining tract tracing and neurochemical markers can provide information on the microenvironment of the origin or termination of distinct pathways



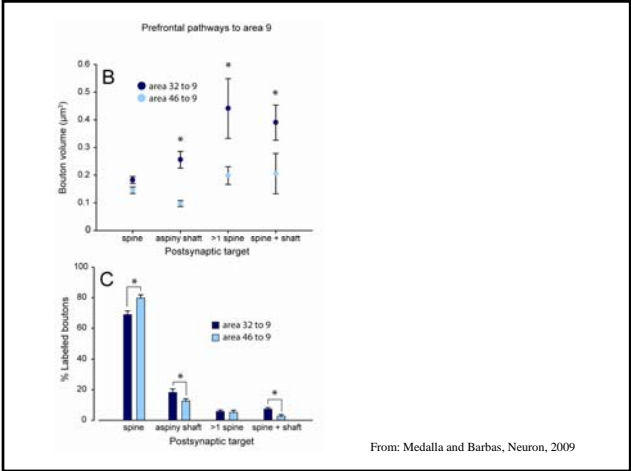
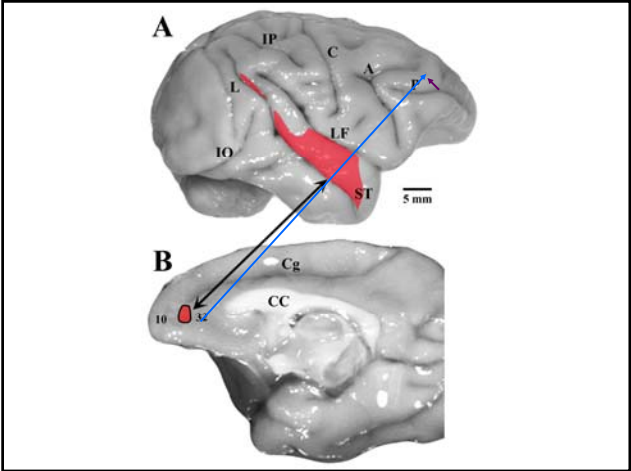
Prefrontal pathways at the synaptic level: axonal boutons terminating in the **middle layers** are larger than boutons terminating in **layer I** of superior temporal auditory association cortex



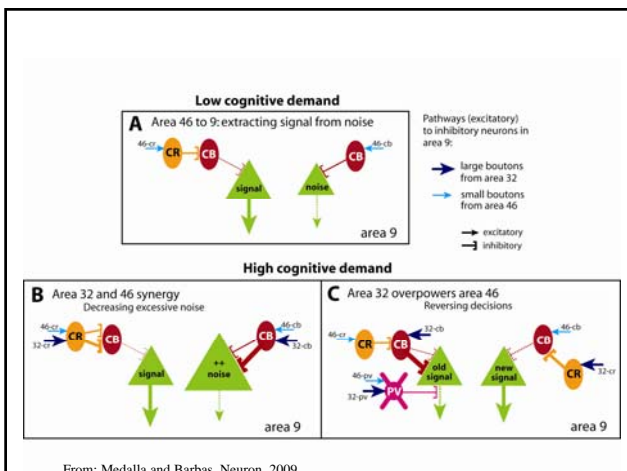
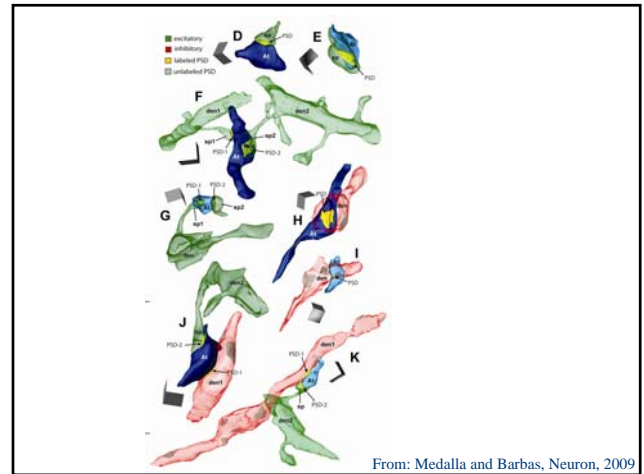
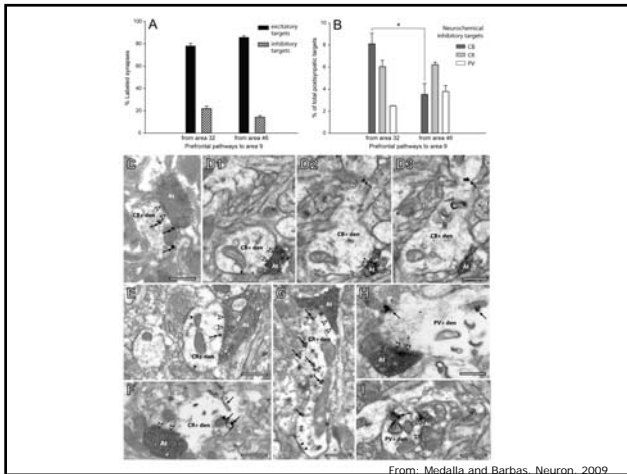
From Germuska et al, Cerebral Cortex, 2006



From: Zikopoulos and Barbas, 2007.



From: Medalla and Barbas, Neuron, 2009



What can pathways tell us about normal function and pathology?

Pathology in schizophrenia: Perspective from pathways

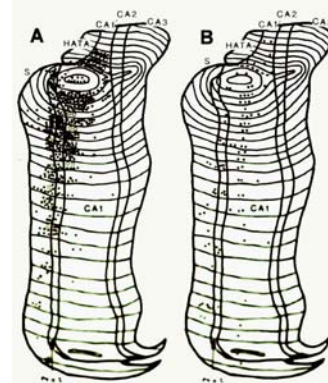
Pathology in schizophrenia

The roots of the disease are in development, affecting the delicate balance of neuronal migration, architecture and ultimately connections

Pathology in schizophrenia affecting cingulate and interconnected structures

Hippocampal formation; the rostral half is affected preferentially in schizophrenia

The rostral half of the hippocampal formation is the principal source of projections to the anterior cingulate



Pathology in schizophrenia

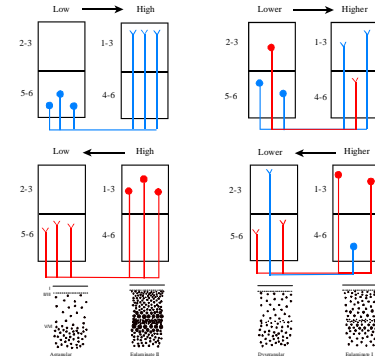
The number of pyramidal (excitatory) neurons is reduced in the deep layers of the anterior cingulate cortex (ACC) in schizophrenia (Benes et al., Biol. Psych., 50, 2001).

The deep layers of ACC project to the upper layers of dorsolateral prefrontal cortex.

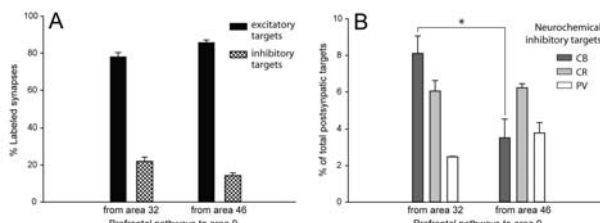
Linking cortical architecture to corticocortical connections

The structural model: Predicting the laminar pattern of connections from cortical structure

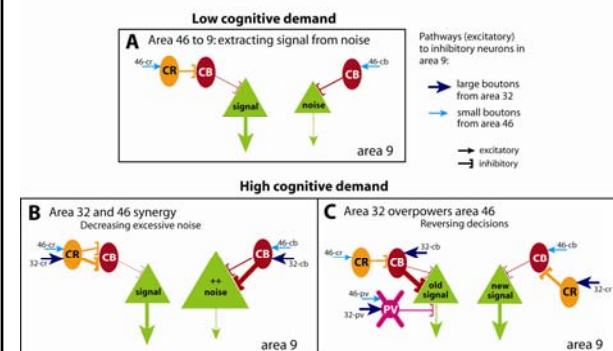
A. Large differences in laminar definition B. Moderate differences in laminar definition



Adapted from: Barbas and Rempel-Clower, 1997



From: Medalla and Barbas, 2009



From: Medalla and Barbas, Neuron, 2009