UNIVERSITY of CALIFORNIA

Exposure Control Plan (ECP)

UC San Diego Environment, Health and Safety Phone : (858) 534 3660 Fax : (858) 534-7982

Name:	me: Rabies Viral Vector				BSL: 2+		
1. Mode of Transmission to Humans (e.g. inhalation, inoculation, mucous membrane exposure, etc.):							
There are replication competent and replication defective or attentuated strains of rabies viral vector at UCSD. The specific strain should be identified on the BUA.							
The most likely sources for exposure of laboratory and animal care personnel are accidental parenteral inoculation, cuts, or needle sticks with contaminated laboratory equipment, bites by infected animals, and exposure of mucous membranes or broken skin to infectious tissue or fluids. Wild type rabies is thought to have caused laboratory infection via exposure to large quantities of aerosolised rabies virus and therefore all aerosolizing procedures with the viral vector must be done in a biosafety cabinet.							
2 Description of the Human Disease associated with this agent or vector (including instances of laboratory acquired infections):							
The wild type rabies virus can cause an acute infection, marked by progressive encephalomyelitis, and is usually fatal. The initial symptoms of rabies resemble those of other systemic viral infections, including fever, headache, malaise, and upper respiratory and gastrointestinal tract disorders. This prodromal phase typically lasts about 4 days, but can last as long as 10 days before specific symptoms develop. Almost all cases of (untreated) clinical rabies are fatal.							
LABORATORY-ACQUIRED INFECTIONS: Two cases of laboratory-acquired wild type rabies infections have been reported and are thought to have been acquired via high concentrations of aerosolized virus across mucous membranes. No cases of laboratory-acquired infections have been reported in the last several decades. There is no documented LAI involving the replication defective rabies viral vector.							
3. Personnel Protection Required (minimum requirement):							
		Laboratory	Vivarium		Laboratory	Vivarium	
Sa	afety Glasses	Χ	X	Shoe Covers		X	
Li	ab Coat	X		Face Shield			
G	loves	X	X	Cap/Head Cover		X	
G	oggles	X	X	Mask			
D	isposal Gowns		X	Other	X		
0	Other - Describe: Betadine must be available in the laboratory to be used in the event of a percutaneous exposure.						
4. Engineering Controls (minimum requirement):							
U	C x	Laboratory	Vivarium		Laboratory	Vivarium	
А	utoclave		Χ	Engineered Sharps Protection			
B	ioSafety Cabinets	X	X	Sharps Containers	X	X	
C R	losed Centrifuge otors	X	X	Closed Centrifuge Tubes	X	X	
0	ther	Х	X				
0	ther - Describe:	A Biosafety	Cabinet must be	e used for all aerosol-generatin	g procedures		

5. Disposal Procedures:

http://blink.ucsd.edu/safety/research-lab/hazardous-waste/medical/dispose.html

Liquids: add bleach to a final concentration of 10% bleach solution (Clorox or other bleach with Cal EPA registration number), let sit for 30 minutes, dispose of in sink (best while running water).

Aspirator flasks: bleach is added to aspirator flasks initially so that the final concentration will be 10%. For tissue culture label the flask as follows: "Tissue culture media disinfected with bleach 9:1." These must be emptied at least weekly.

Solids: disposed of in labeled, double red biohazard bags held in labeled, rigid, covered containers. Transport to biohazard collection area in a closed rigid container for final disposal by EH&S. These must be emptied at least weekly.

6. Disinfection Procedures:

Surface: 10% household bleach solution made up fresh daily (Clorox or other bleach with Cal EPA registration number) allowing 5minutes contact time.

OR

Surface: 75 ppm of CA EPA approved iodophor (e.g. Wescodyne) allowing 5-minutes contact time.

Biosafety cabinets can be disinfected first and then cleaned with ethanol to prevent corrosion.

7. Recommended/Required Vaccinations or other Medical Surveillance:

Prior to working with the rabies viral vector, all personnel are required to be counseled through the campus Occupational Health Nurse or Center for Occupational and Environmental Medicine (COEM), which includes completion of the Rabies vaccine accept/ decline form. Immunization is offered for individuals at high risk for exposure.

8. Employee Exposures- first aid procedures:

a. Eye exposure from splash or aerosols - rinse a minimum of 15 minutes in eye wash or flush area with water.

b. Skin exposure - wash area with soap and water for 15 minutes, then apply povidine iodine (betadine)

c. Needle stick and/or sharps exposure - wash wound area with soap and water for 15 minutes, then apply povidine iodine (betadine)

d. Contamination of clothing - remove the contaminated clothing and place in biohazard bag, shower with the emergency douse shower, and put on clean clothes.

e. Spill or release - Monday through Friday, 8a - 4:30p call EH&S; after hours call Campus Police

9. Employee Exposure - seek medical follow-up from the following medical providers: (TAKE THIS ECP WITH YOU) 24-hour walk-in service:

Hospital Emergency Room Or nearest Medical Center Emergency Room

Call your campus Occupational Health contact to determine if a follow up visit is required. Most exposures will require a visit to Occupational Health.

Monday - Friday, 8a - 4:30p

Post-exposure rabies prophylaxis with HDCV or PCECV together with the administration of rabies immunoglobulin (RIG) is highly effective, although this should not be used in persons who have previously received complete vaccine regimens (pre-exposure vaccination) who require vaccination only

10. Report All Injuries, Illnesses, and Exposures to EH&S:

Report the injury incident to their immediate supervisor and complete your campus injury report form. Link below: http://blink.ucsd.edu/safety/occupational/reporting.html

11. Required Biosafety Training:

Viral Vector Training is required before work with this agent begins.

Required campus safety training found on UCSD Blink: http://blink.ucsd.edu/safety/resources/training/general-lab.html#Principal-Investigators

Laboratory specific training on hazards, exposure evaluations, and the required precautions for experimental procedures used with this agent need to be provided by Principal Investigator.

12. Lab specific instructions:

For this particular experiment, Amphyl will be used as the disinfectant instead of Chlorine. The amount used will be 1:200 Amphyl. Goggles to be used by personnel.