

## **VIRAL DECONTAMINATION FOLLOWING INJECTION SURGERY (SOP-70)**

Prepare two solutions:

- 1:200 Amphyl
- 70 % (v/v) EtOH in H<sub>2</sub>O

Personal protection equipment for this includes hair cover, goggles, double nitrile gloves, a disposable gown, and shoe covers over closed toed shoes.

Excess virus will be inactivated with 1:200 Amphyl in the Biosafety cabinet and discarded as chemical hazardous waste.

All work surfaces will be decontaminated by wiping with 1:200 Amphyl for a period of at least five minutes followed by clean-up in 70 % EtOH.

All equipment used in the experiment, including but not limited to stereotaxic holder and electrode manipulator, Nanoinjector, surgical tools, physiological headstage (if used), perfusion needles and lines, and viral transport Dewar will be decontaminated by washing in 1:200 Amphyl for at least five minutes followed by a wash in 70 % EtOH for purposes of sterility.

All disposable items will be decontaminated with 1:200 Amphyl and disposed of in biohazardous waste containers.

### **For the case of replication competent viruses**

- We will use existing cages and place the cages in a red Biosafety bag and notify ACP for the bag to be autoclaved.
- Any animal that is found morbid prior to perfusion shall be sealed in a plastic screw cap container filled with 1:200 Amphyl. After 10 minutes the liquid is poured off and the carcass disposed of in double clear colorless bag and placed in the ACP carcass disposal cold box.
- If a cage change is required, it is our responsibility to do so.
- All dirty cages will be placed into red biohazard bags that are sprayed with 1:200 Amphyl on the outside and left in the room for ACP to autoclave.